

Glucosinolates and sulphur-rich cells

Distribution in roots of field-grown canola

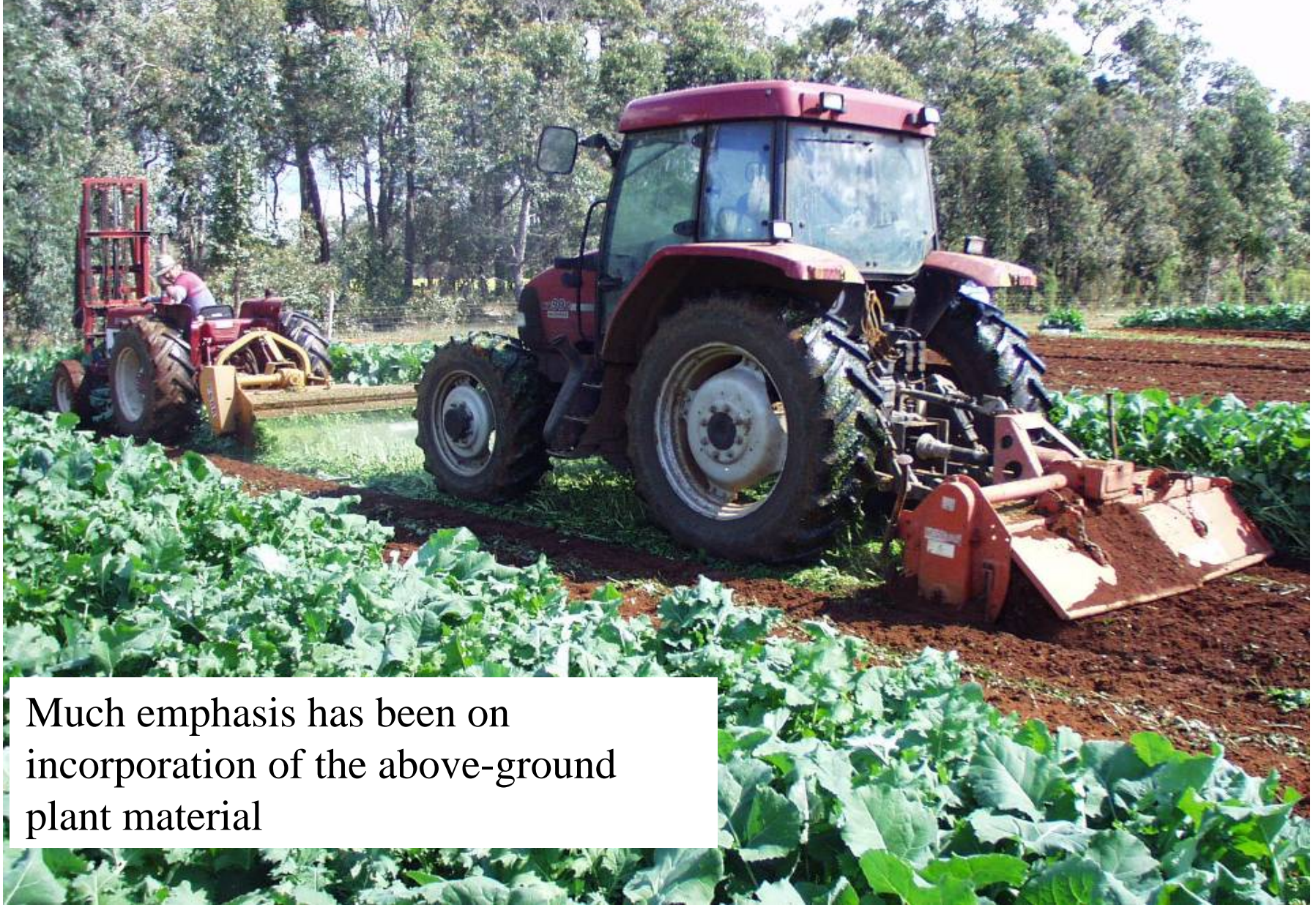
Margaret McCully, Celia Miller, Susan Sprague,
John Kirkegaard



CSIRO Canberra

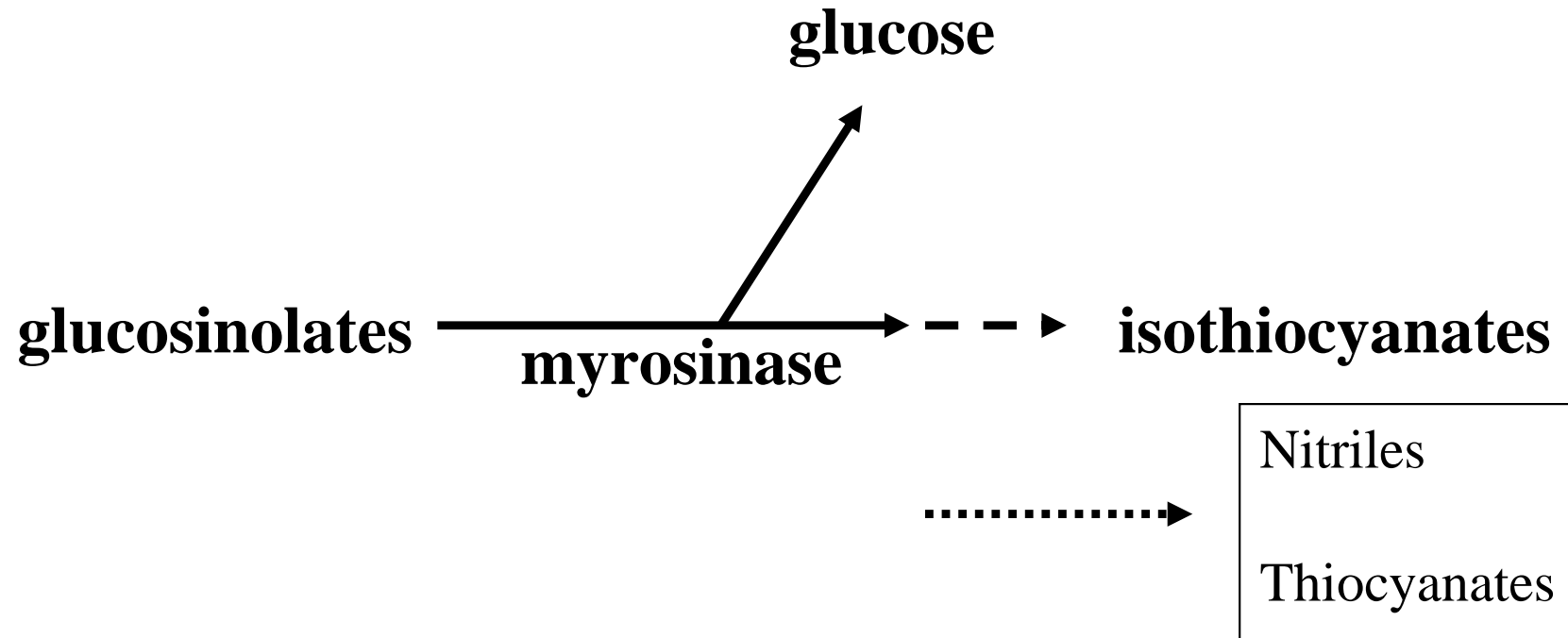


Biofumigation uses residues of glucosinolate-containing plants for pest and disease control



Much emphasis has been on incorporation of the above-ground plant material

Biofumigation harnesses the
'mustard bomb' reaction



Why focus on *Brassica* roots?

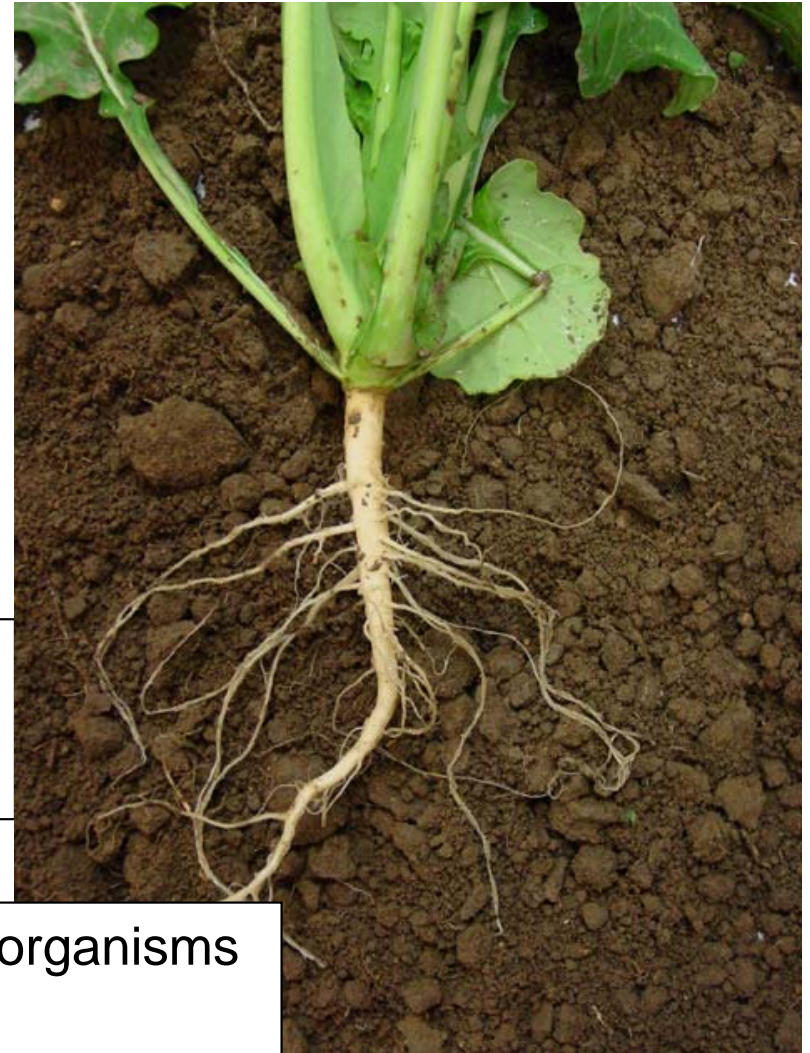
On average roots have 4.5 X
GSL concentrations in shoots

Van Dam *et al.* 2008

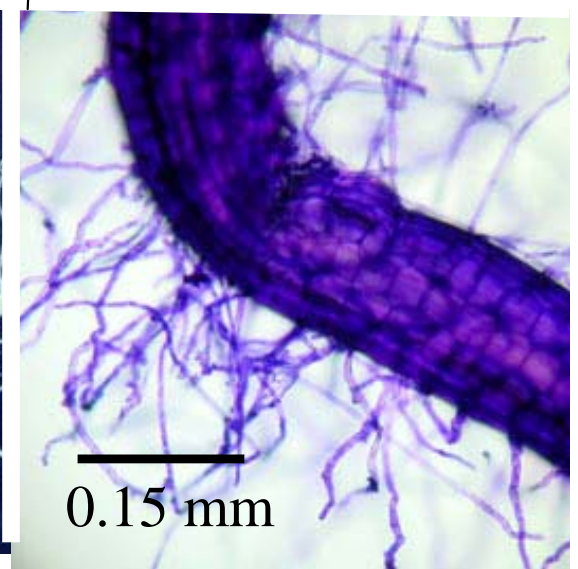
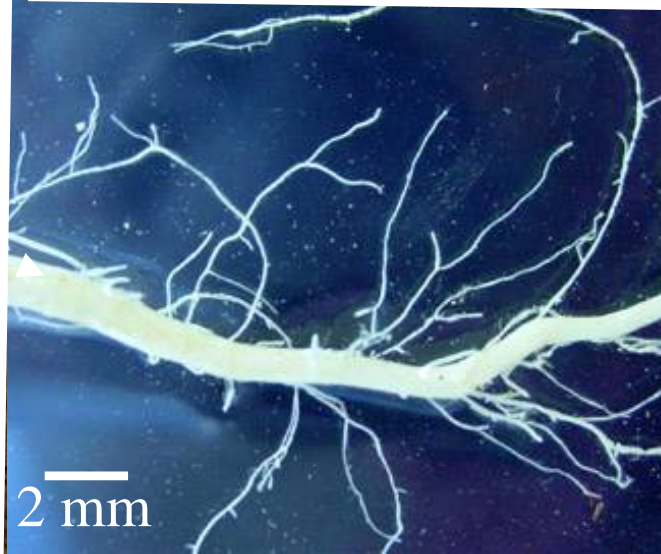
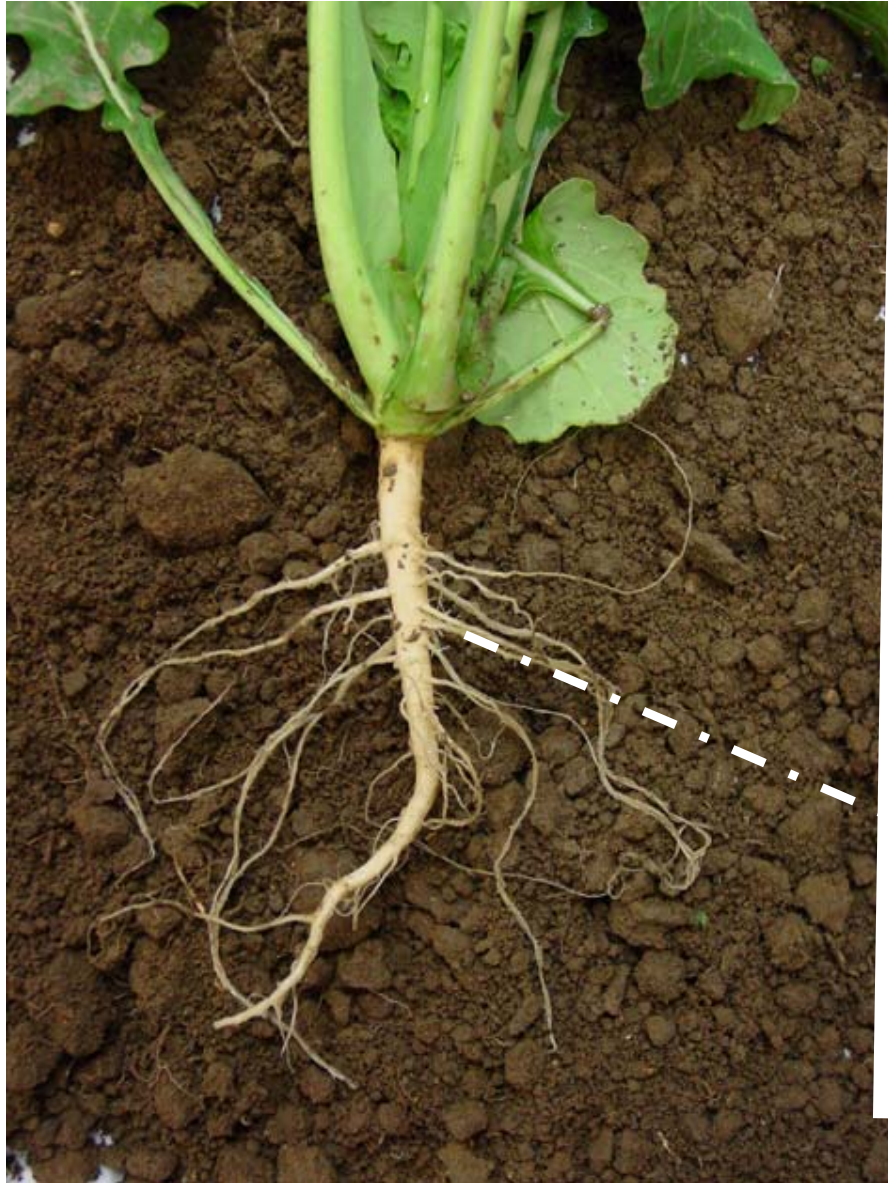
2-PE glucosinolate predominates in
roots. Little or none in shoots

2-PE ITC the most toxic to soil organisms
and most herbicidal

Roots are continuously exposed to potential pathogens
and predators



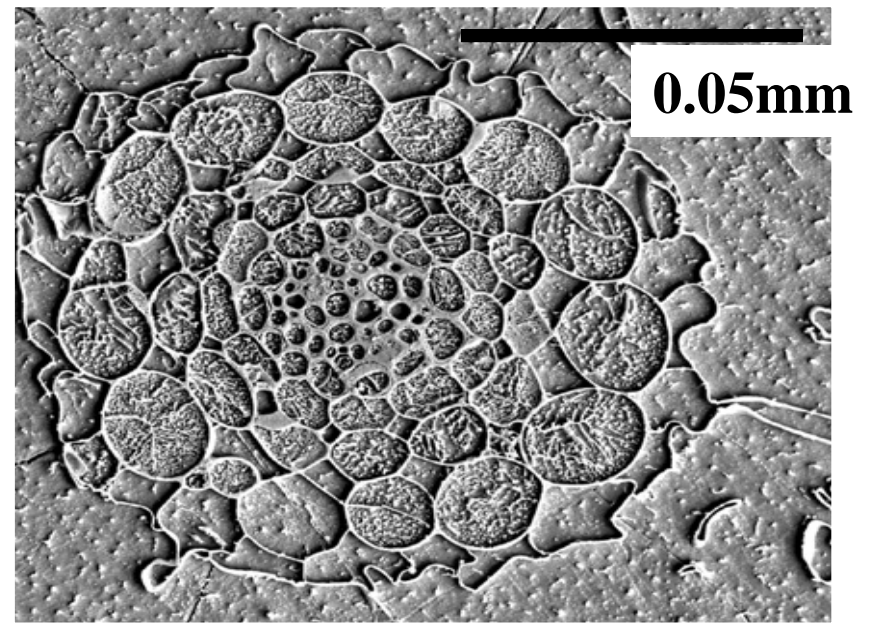
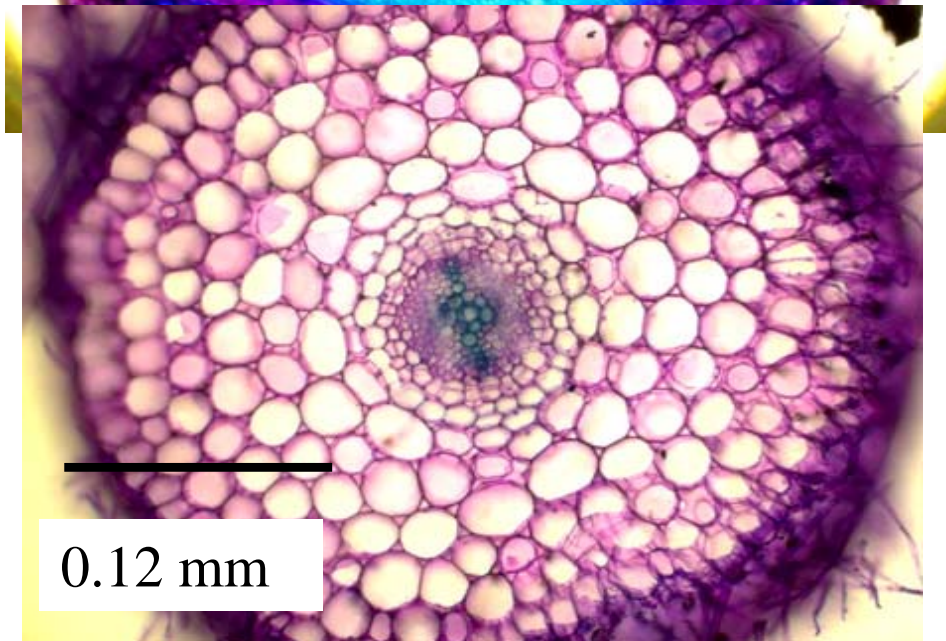
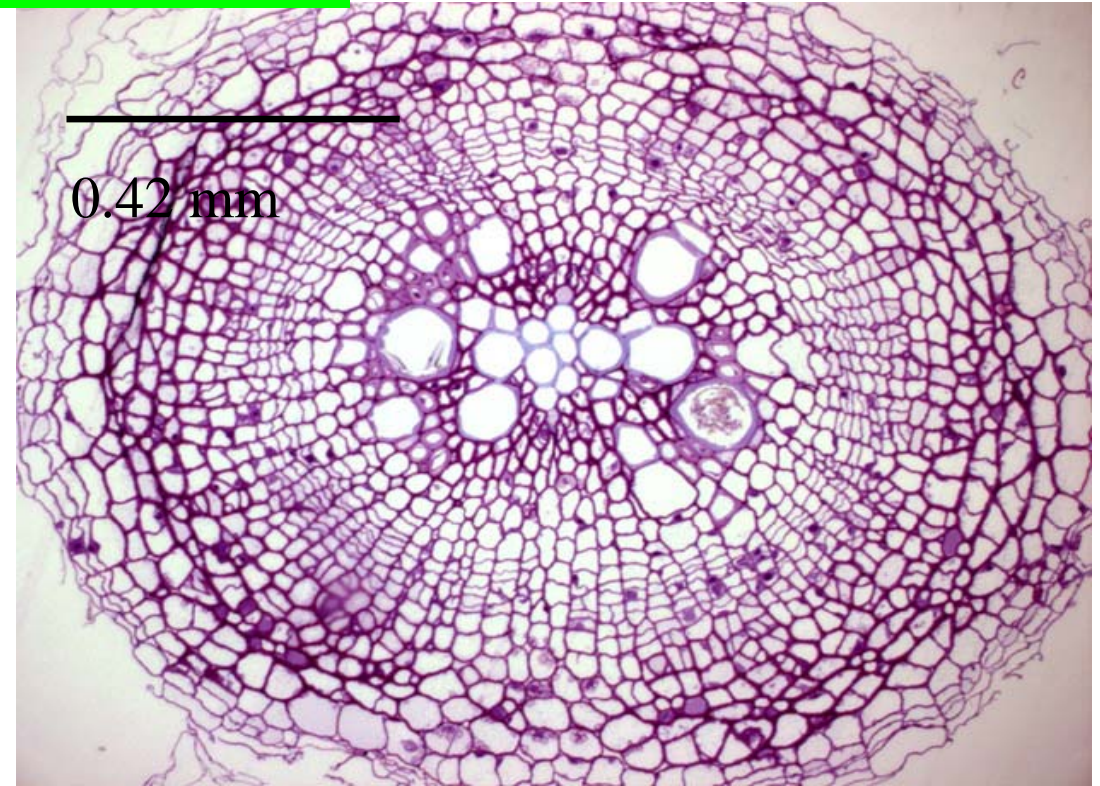
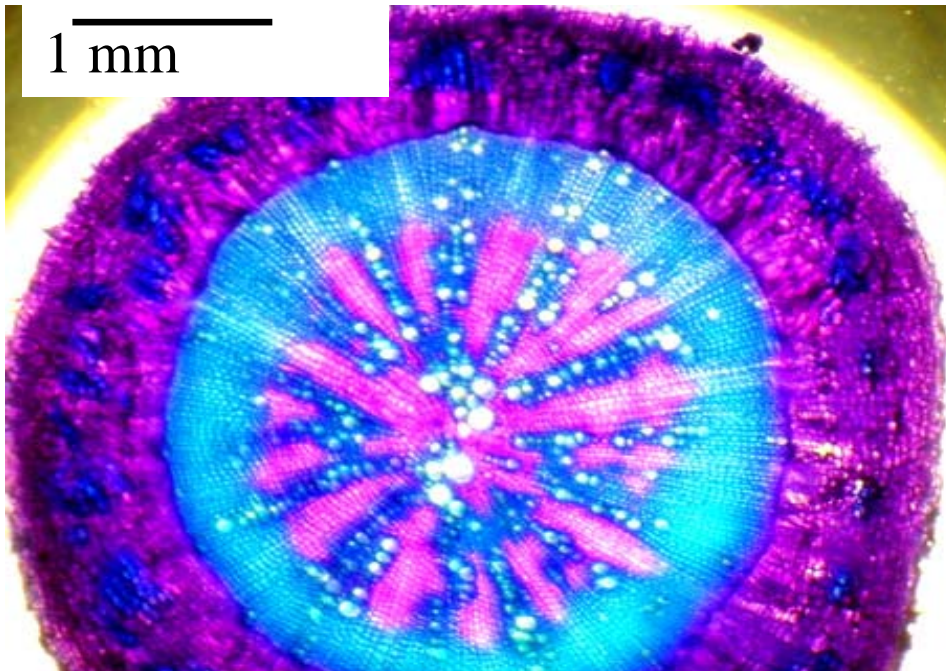
Canola root system hierarchy



The very fine roots are short-lived and proliferate as plants mature



Component root anatomy



Localizing glucosinolates in roots

In particular root types

In different tissue regions of each root type

Cell-specific localization



GSL in particular root types

Varying GSL concentrations in different root types

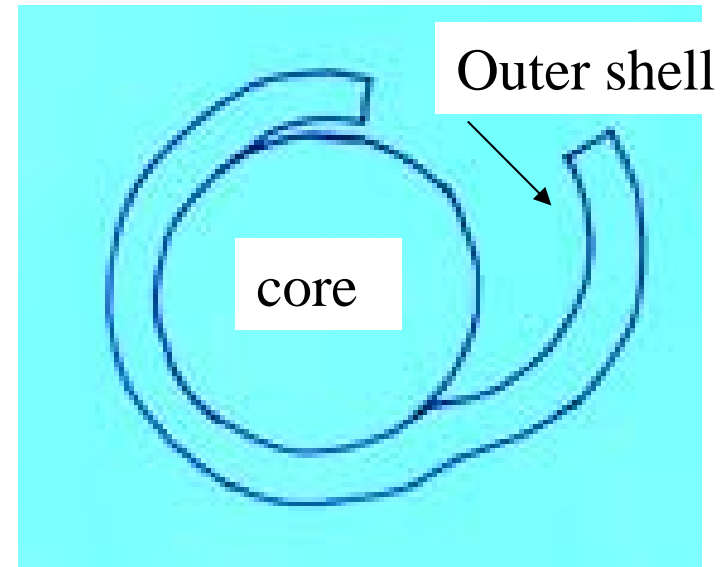
	Large branches >2 mm	Tap roots	Small branches < 2 mm
Concentration ($\mu\text{mol g}_{\text{dw}}^{-1}$)	15	7	7
Contribution to root system (%)	52%	35%	13%

Kirkegaard & Sarwar 1999

GSL localization in two tissue regions



Periderm	↑	2nd Xylem
Pro. Pericycle		
2 nd Phloem		



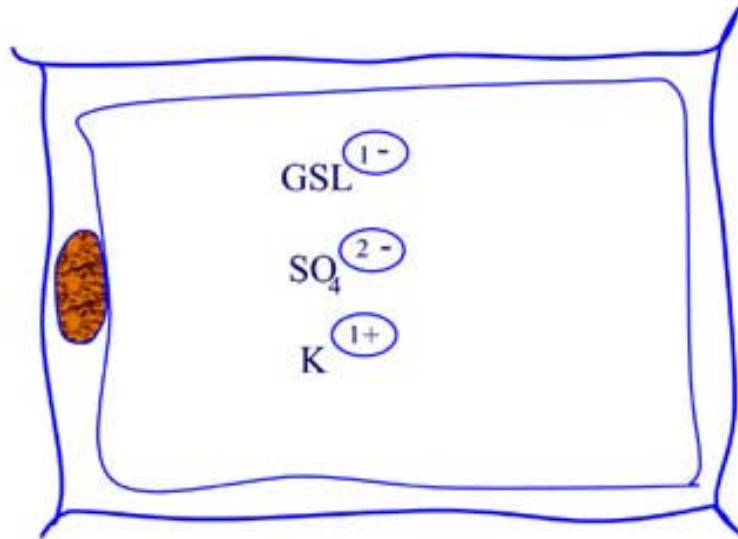
Freeze-dried roots with secondary thickening split easily in the cambial region into an inner core and outer shell

Ground pieces analysed for 2-PE GSL by HPLC

2-PE-glucosinolate content of canola root tissue regions

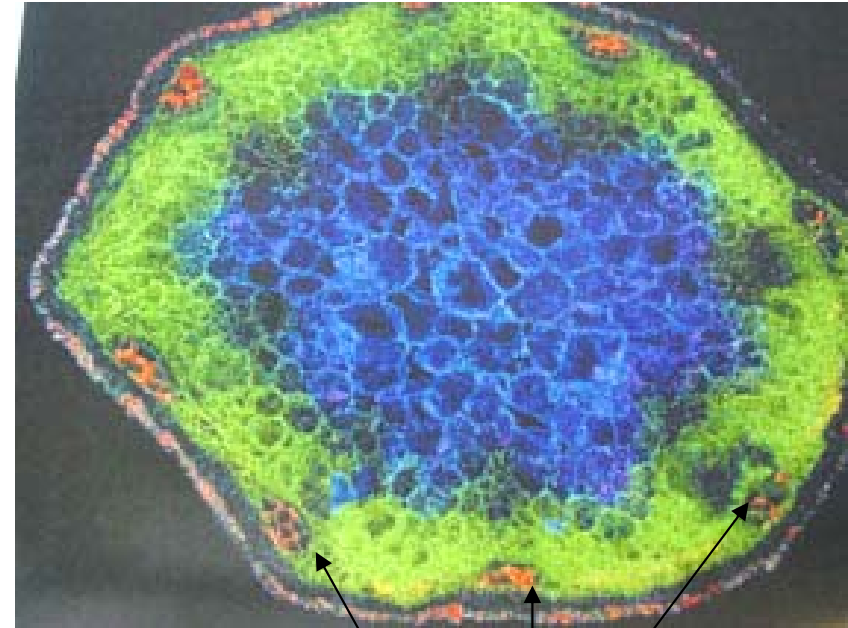
Root type	Tissue region	Percent total root system dw n =20	Glucosinolate concentration	
			$\mu\text{mol g}^{-1} \text{ dw}$	mM
Hypocotyl/ transition region	Outer	14.2	11	2
	Core	25.3	5.5	1
Rest of tap root	Outer	24.9	14	2.5
	Core	29.2	6.4	1
Large branch roots	Outer	2.6	25	4.4
	Core	1.3	5	0.9
Small branch roots	Entire	2.6	16	3

Cell-specific localization of GSL



GSL in cell vacuoles

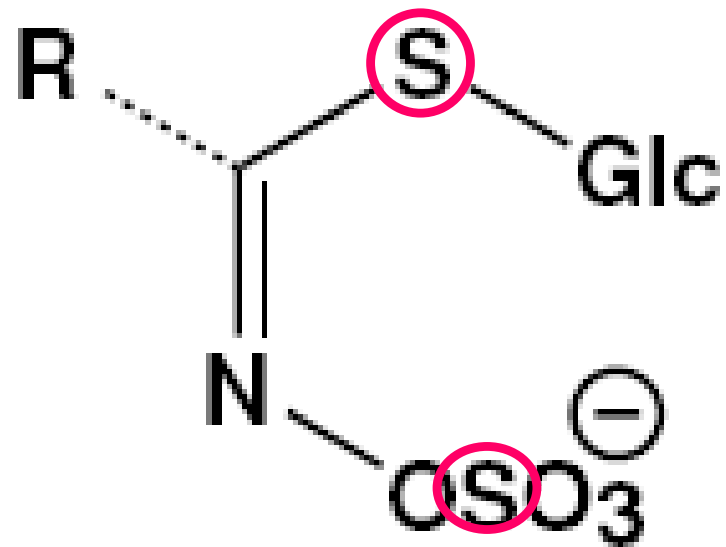
GSLs are water soluble so are only retained *in situ* by cryo-fixation



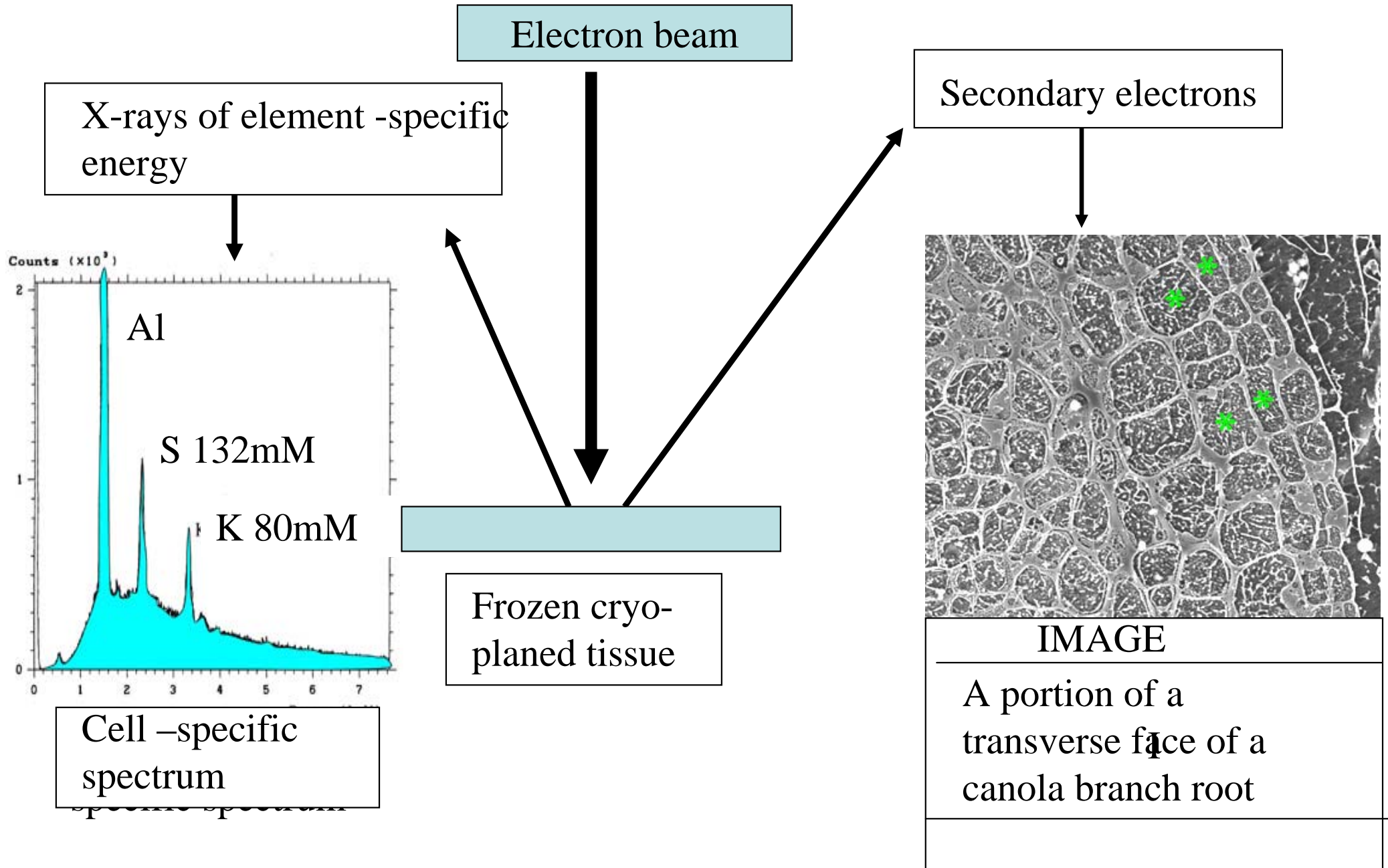
GSL localized in S cells in Arabidopsis flower stalks

Koroleva *et al*
(2000)

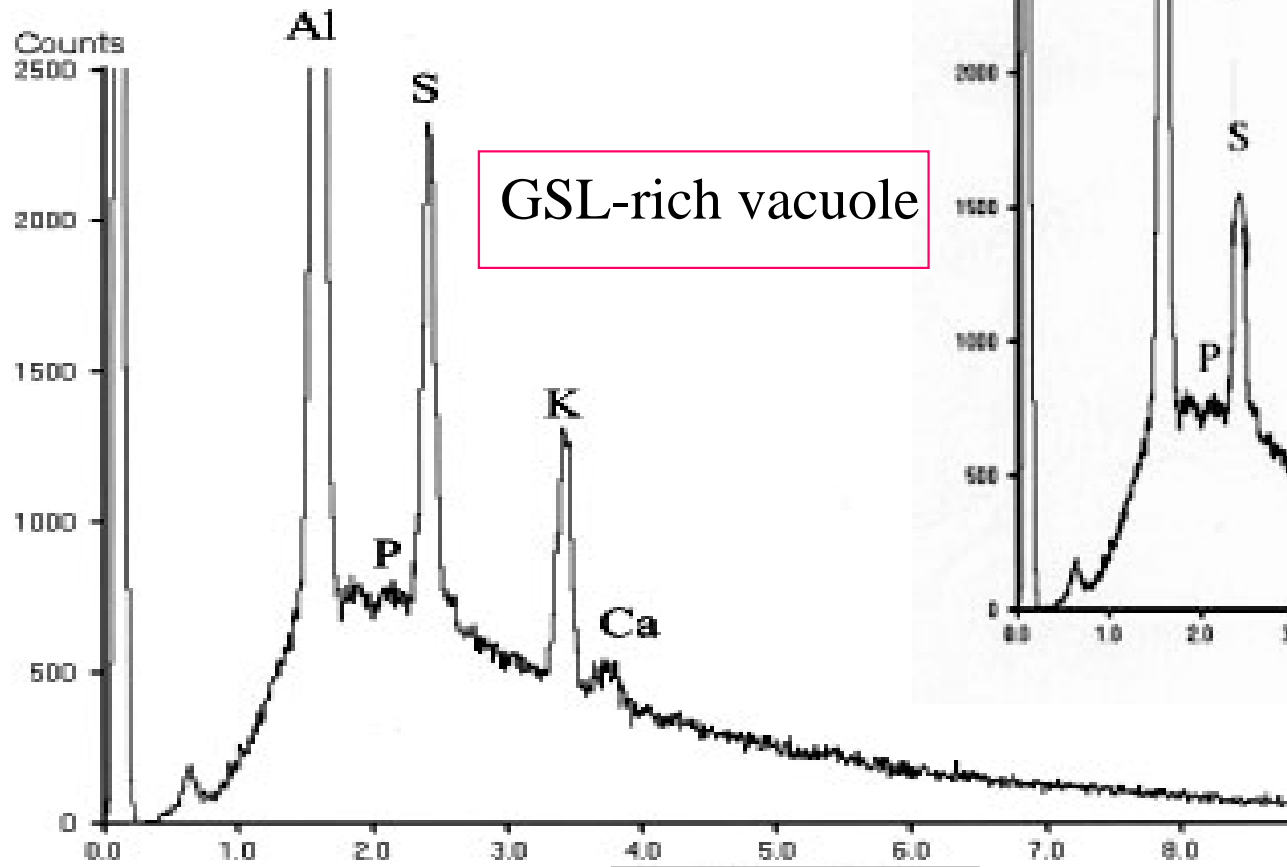
Glucosinolate structure



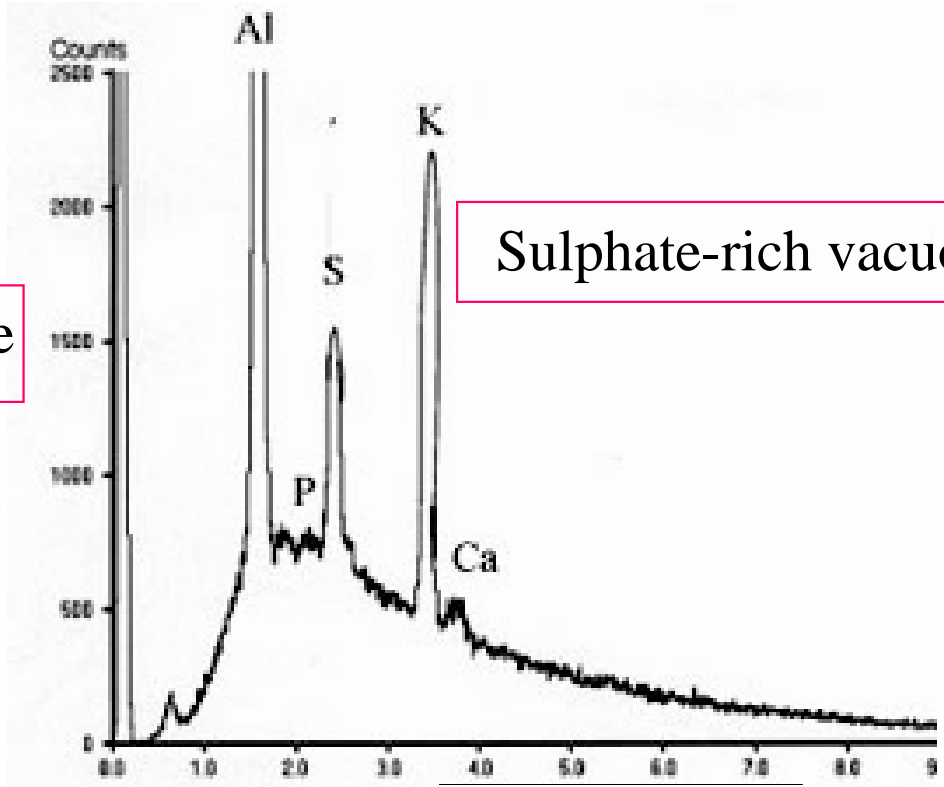
Cryo-Analytical SEM



Vacuoles in GSL-rich cells have high S/K ratios



GSL-rich vacuole

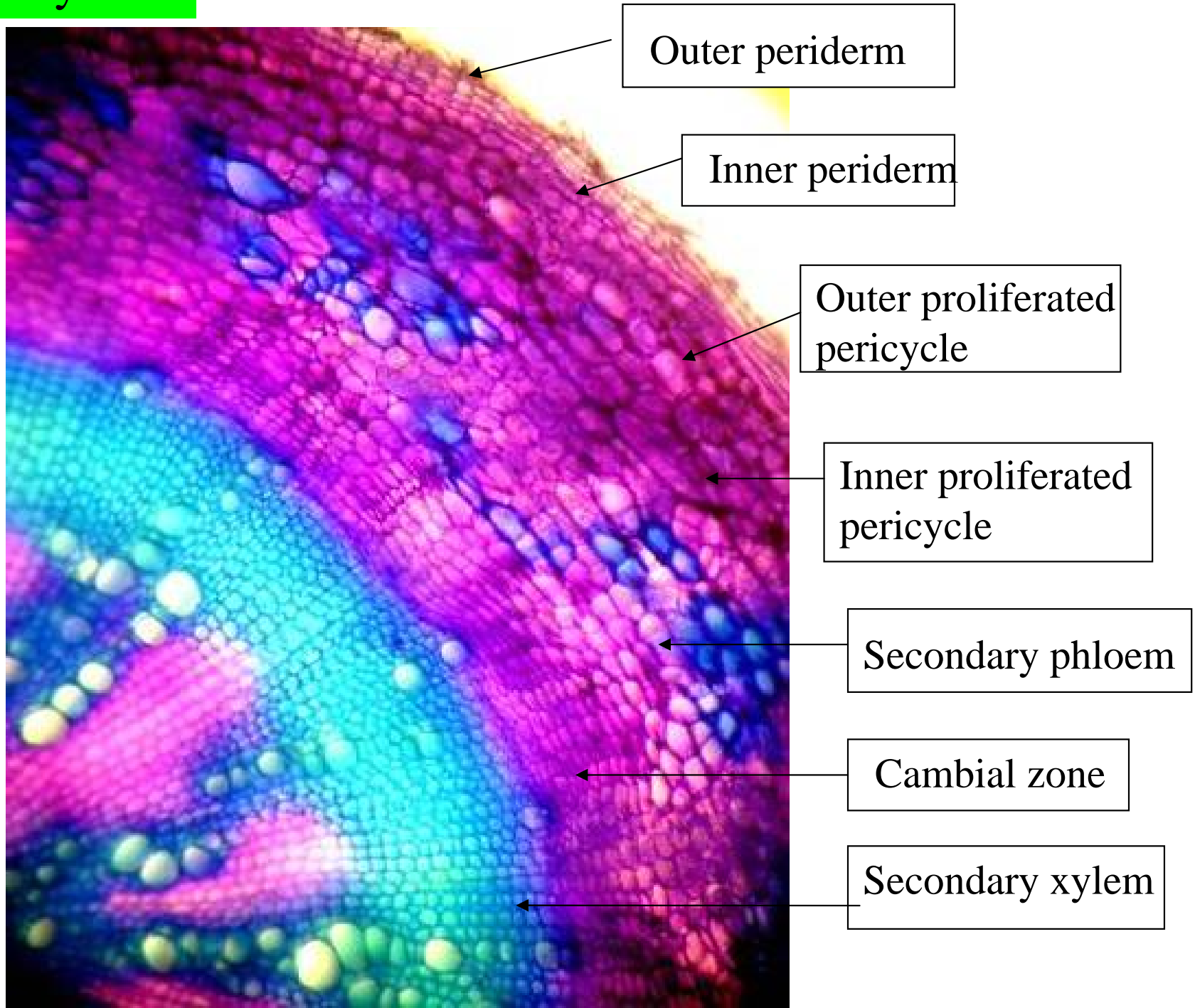


Sulphate-rich vacuole

X-ray energy

X-ray energy

Tissues analysed



Using [S] to determine GSL
concentration

$$[\text{GSL mM}] = \frac{[\text{S mM}] - 50}{2}$$

GSL a monovalent anion balanced by 1K⁺

Sulphate a divalent anion balanced by 2K⁺

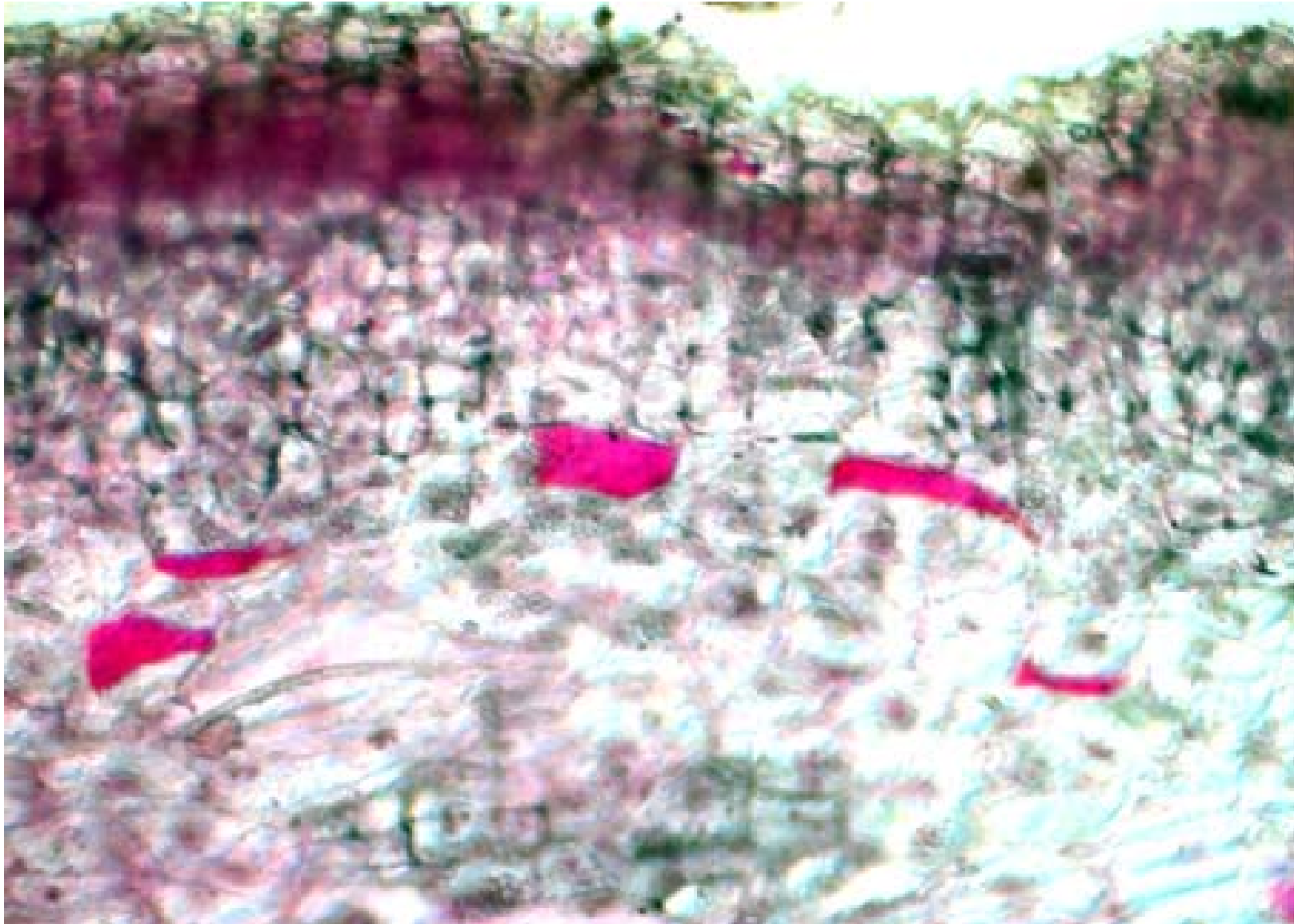
Glucosinolate concentrations in Secondary Tissues

% of cells analysed

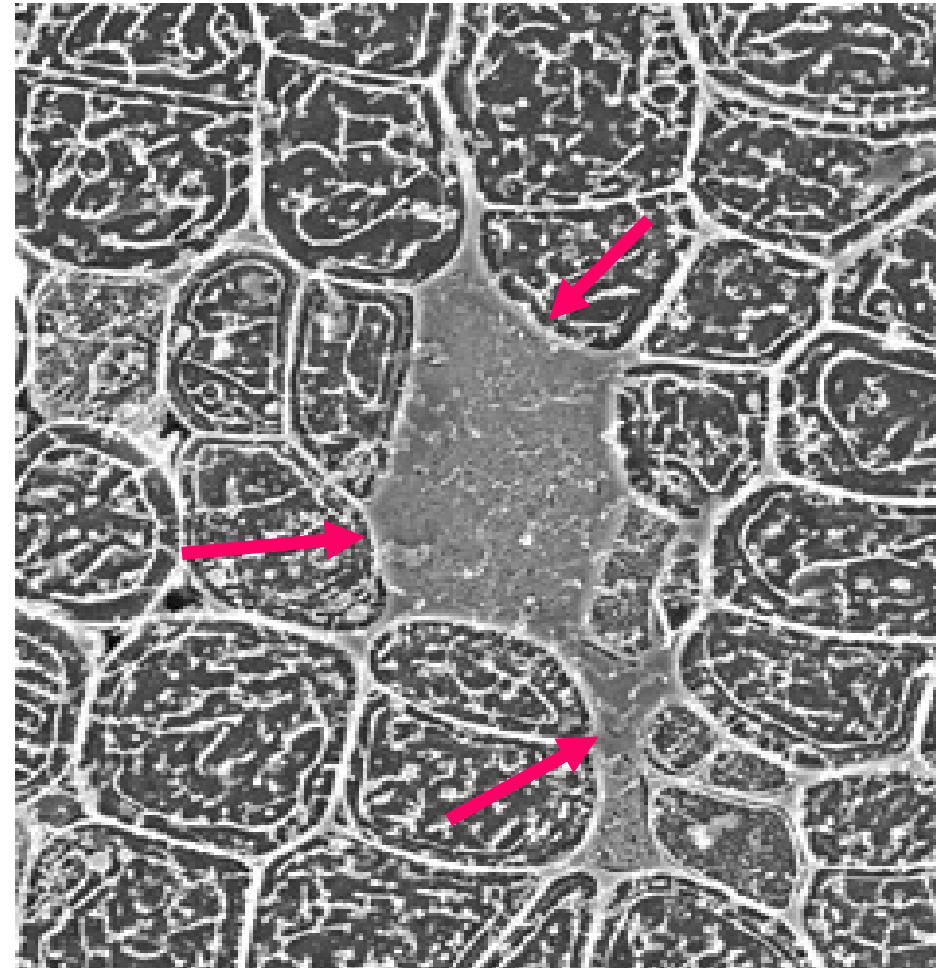
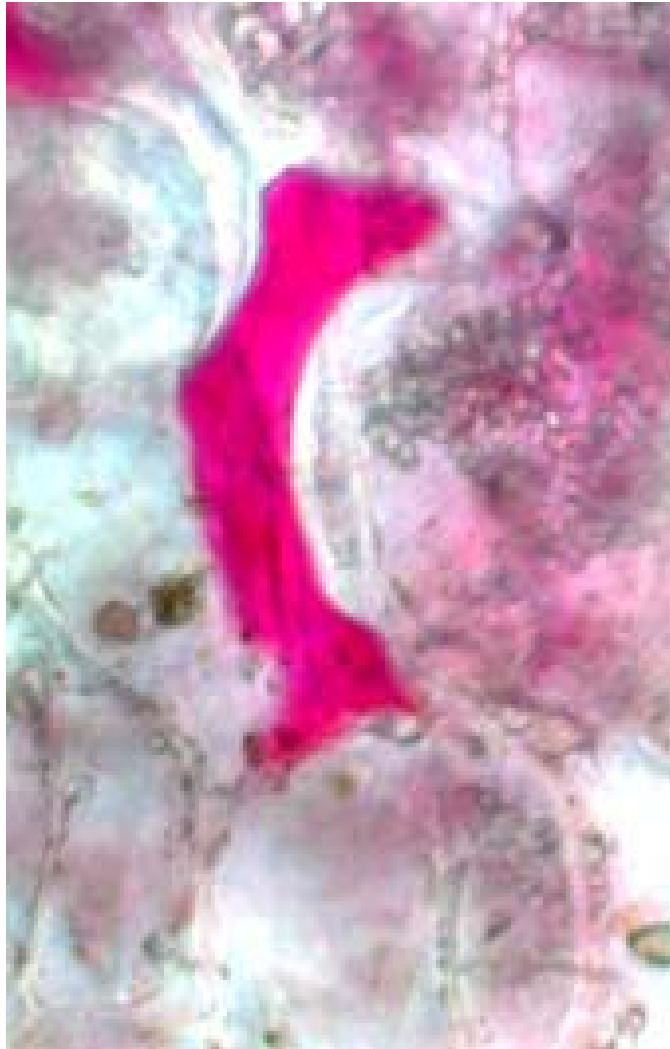
Tissues	>25 mM	>50 mM	>75 mM	>100 mM	N (roots)
Outer periderm	20	7	1	0	67 (20)
Inner periderm	66	46	22	9	94 (20)
Outer pericycle	15	3	0	0	67 (20)
Inner pericycle	2	1	0	0	166 (21)
Remaining 2 ^{ry} tissues	0	0	0	0	156 (24)

McCully et al. 2008

Myrosin cells identified by their irregular shape and strong staining for protein

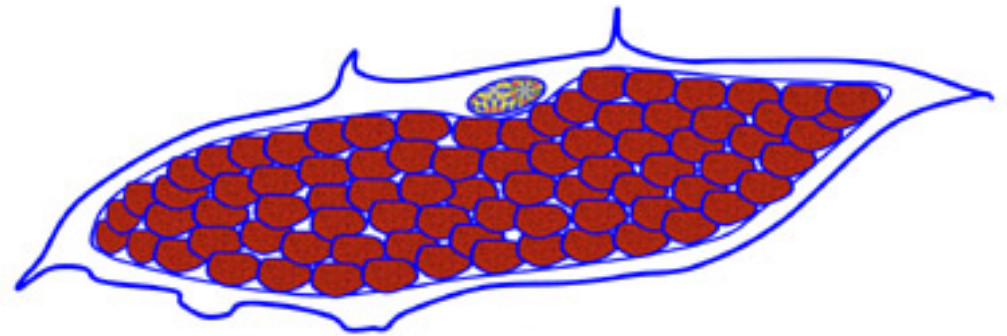


Myrosin cells easily located in the cryo-SEM



Myrosin cells

Are there GSLs in myrosin cells?



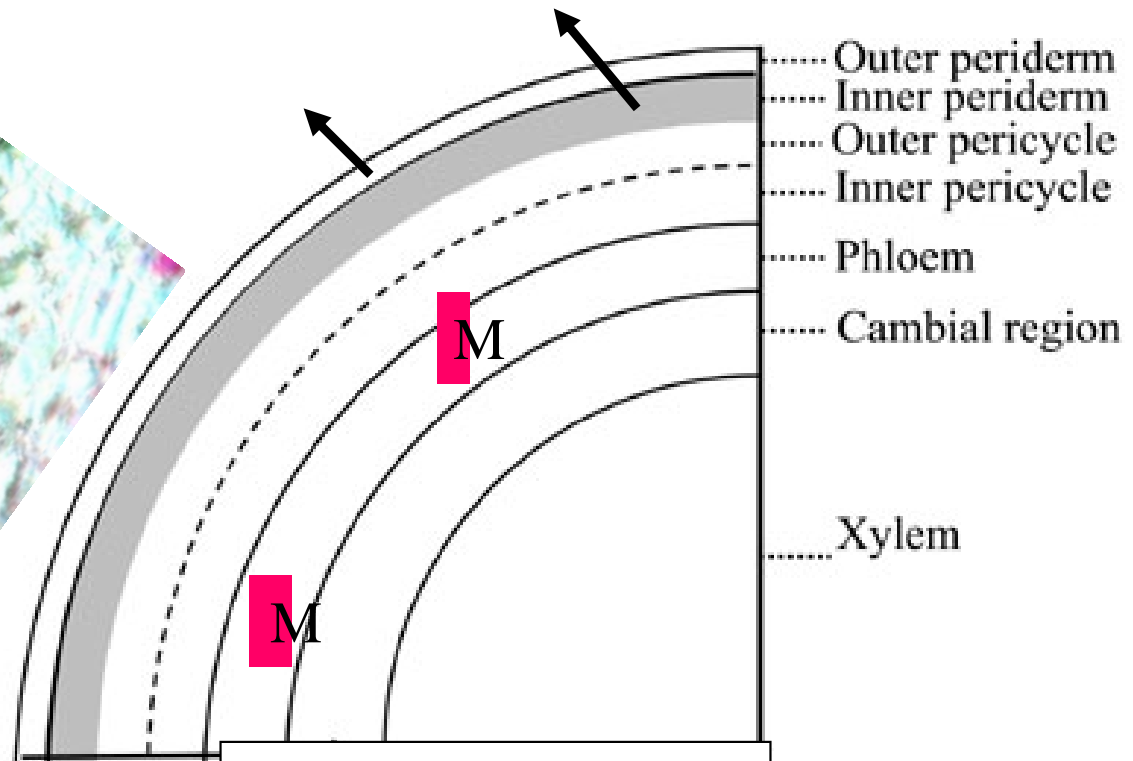
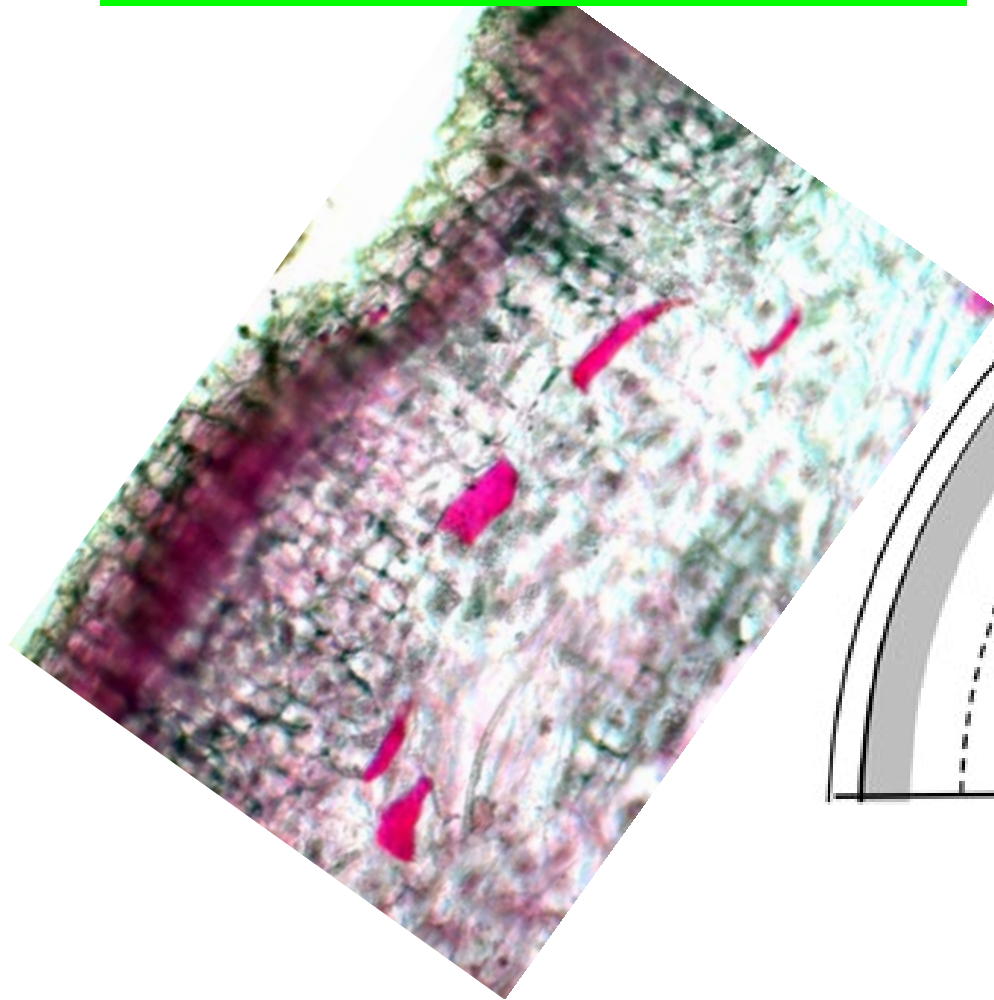
Myrosin cell

	# cells analysed	Mean [S]	Mean [K]
Myrosin cells	8	20 mM	72mM
Adjoining cells	20	32 mM	72 mM

S/K ratio for both cell types = 0.2

How does the “mustard bomb” work in undamaged growing roots?

GSL-rich periderm cells are shed and replaced as root increases in diameter



Myrosin cells remain in situ ?

Isothiocyanates in the rhizosphere ?

2 PE ITC in canola rhizospheres

Highest concentrations at bolting or flowering

Concentrations variable : 0 – 12 nmol g_{dw}⁻¹

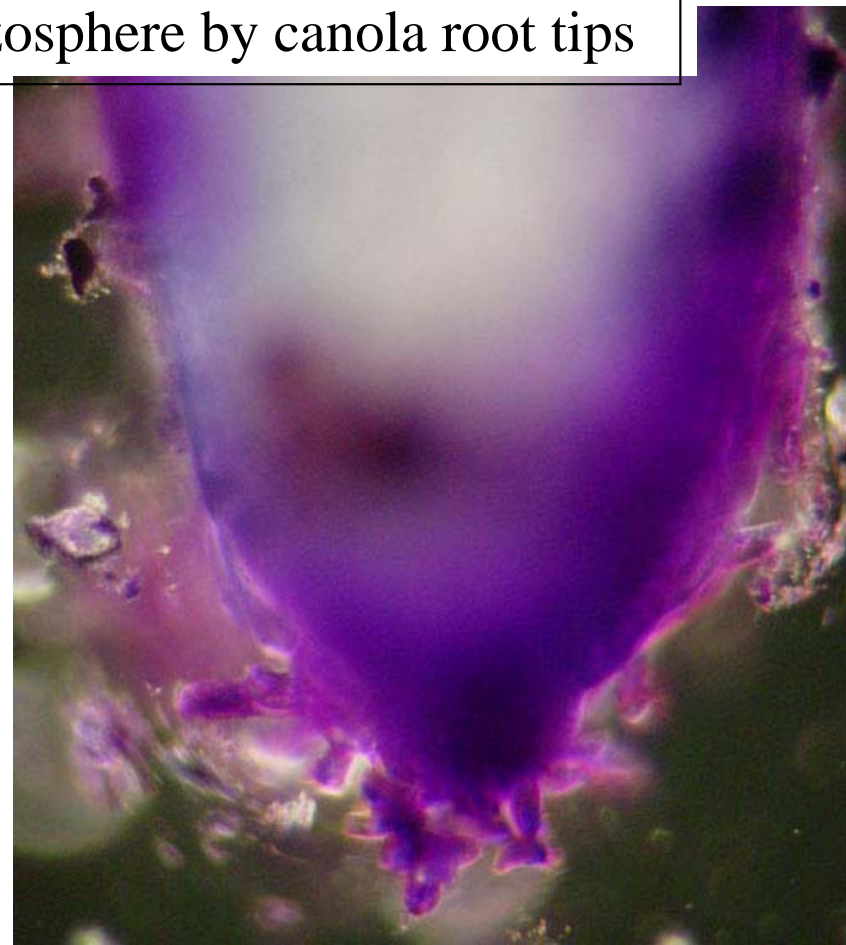
Concentrations highest next to large branch roots

Rumberger & Marschner (2003, 2004)

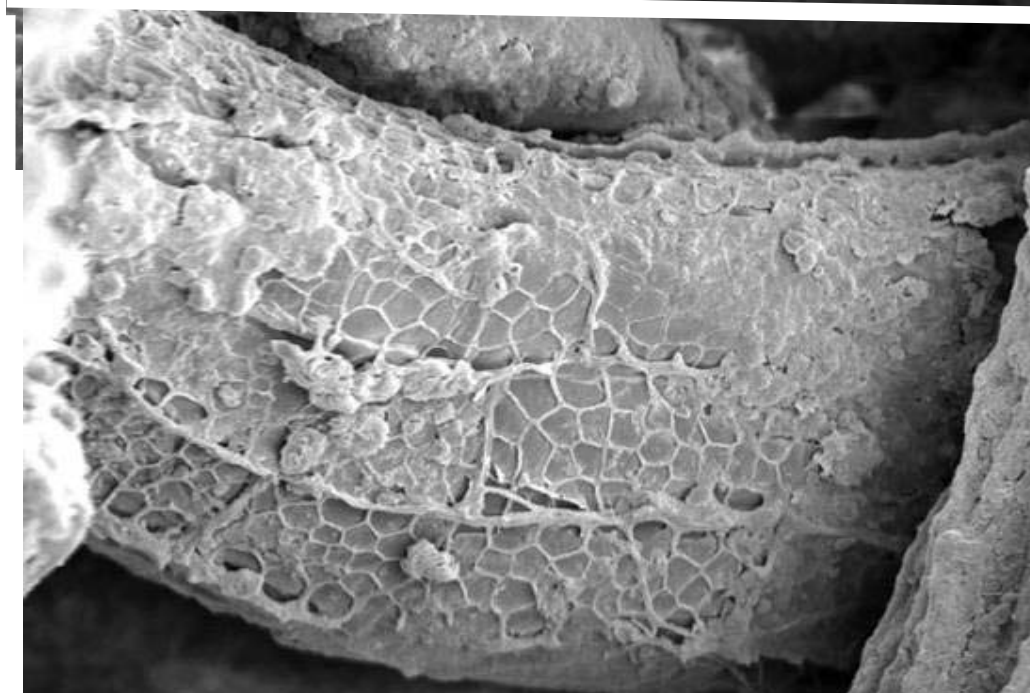
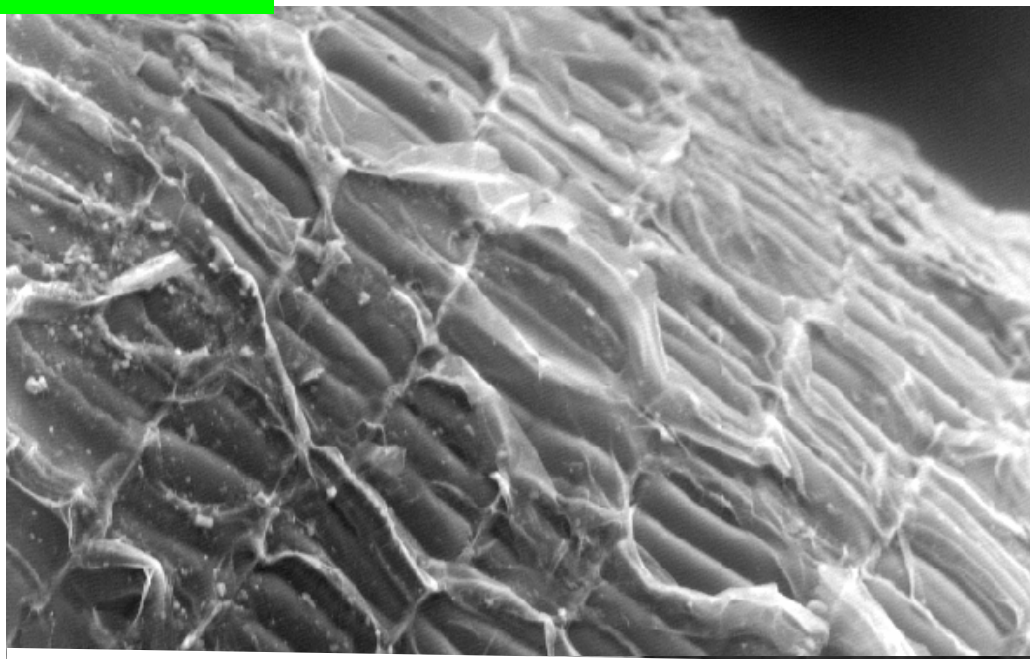
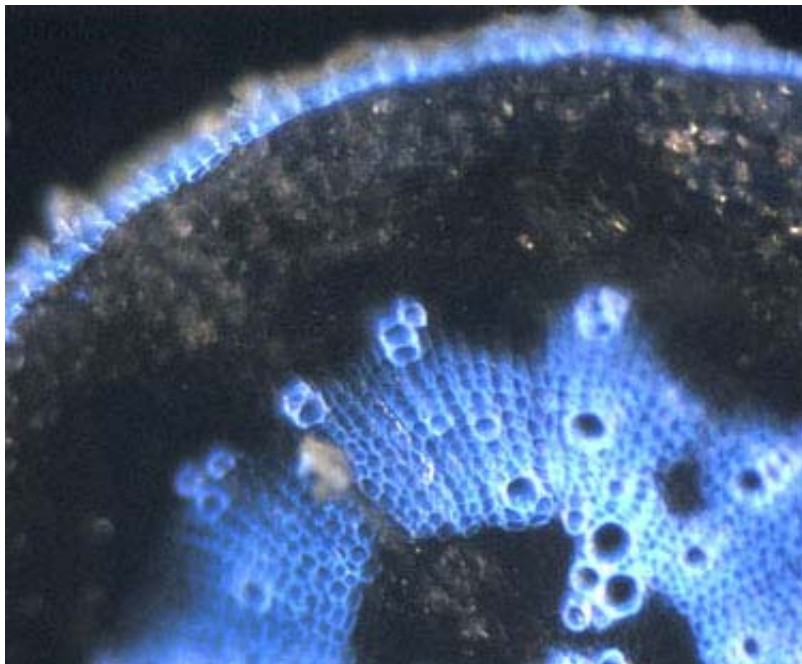
Does myrosinase persist in the rhizosphere?

Myrosinase in the rhizosphere has been reported e.g. Borek *et al.* (1996): Al-Turki & Dick (2003)

Mucilage release to the rhizosphere by canola root tips



Do aliphatic-rich wall residues from periderm cells adsorb 2-PE ITC?



Self protection; Local biofumigation

On average roots have 4.5 X
GSL concentrations in shoots

Van Dam *et al.* 2008

2-PE glucosinolate predominates in
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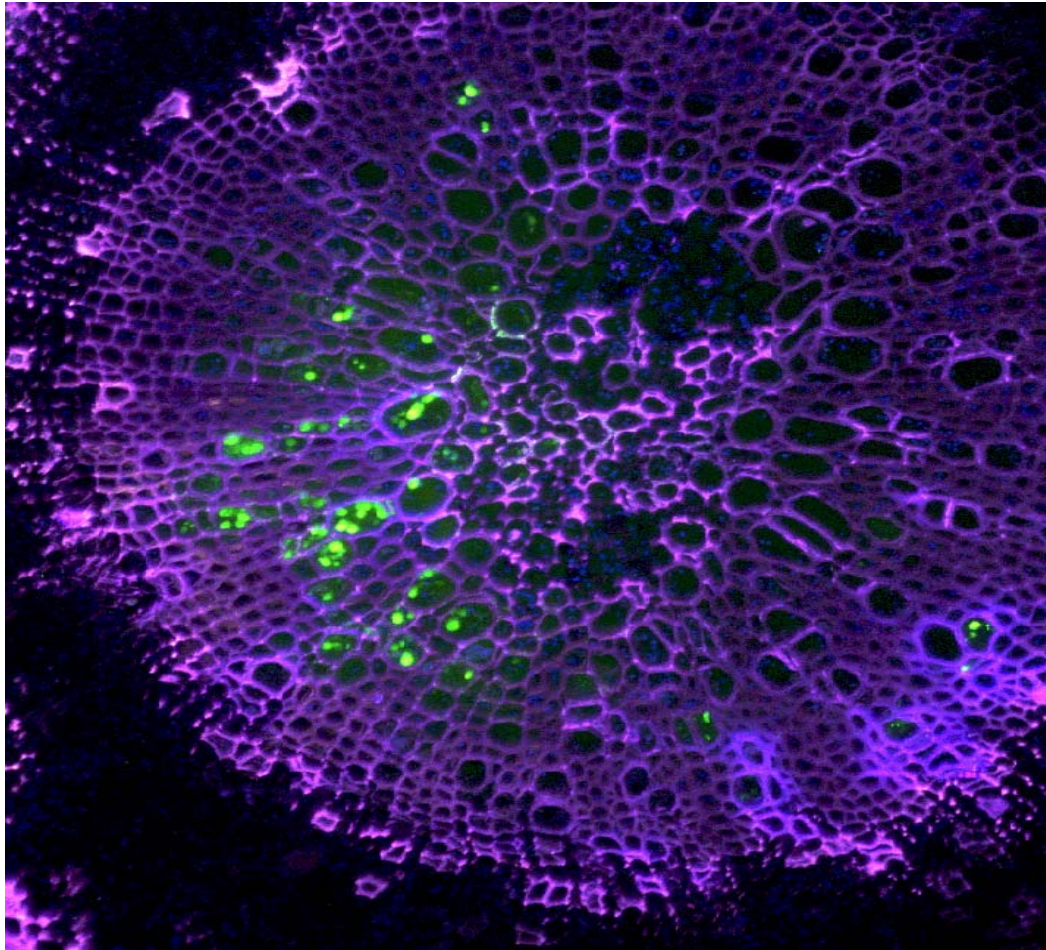


2-PE ITC the most toxic to soil organisms
and most herbicidal

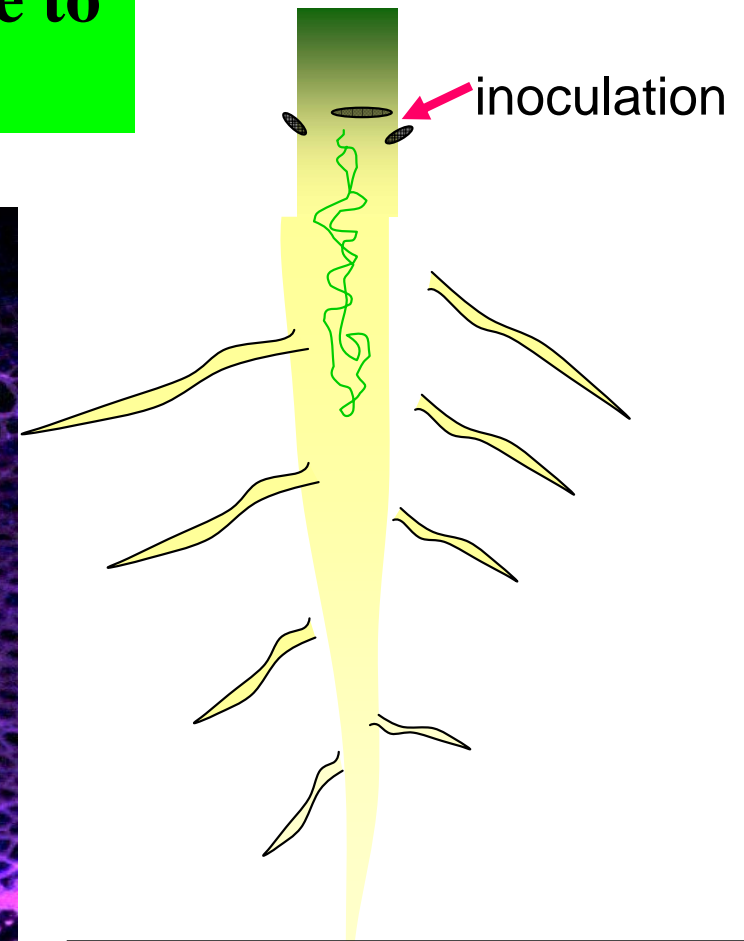
Roots are continuously exposed to potential pathogens
and predators



Does peripheral GSL positioning help confine *Leptosphaeria maculens* hyphae to main root core till post-flowering?



Susan Sprague 2007



GFP-expressing hyphae grow downward into the root secondary xylem core